Effects of In-Ovo Injection of sialic acid on Chick's embryonic development and physiological traits

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Abstract: This study was conducted at the College of Agriculture – University of Anbar, Iraq. From 16 January to 5 February 2022, this study aimed to investigate the effect of injected egg hatching at different sialic acid times in growth and embryonic development. Four hundred eggs of hatching types (Ross 308) were injected with different sialic acid concentrations at 0 days (before placing in the incubator), 7 and 14 days of incubation. Eggs were divided into four groups (100 eggs each) as follows: T1: The control group was placed in the incubator without injection. T2: Injected with a dose of 100 µg sialic acid at the age of zero. T3: Injected with 100 µg sialic acid dose at 7 days of incubation. T4: Injected with 100 µg sialic acid dose at 14 days of incubation. Statistical analysis was performed (CRD) (P=0.05); results show: Increase in the e length of the embryo, the diameter of the vascular region and the number of pairs of somites at 3 days of incubation for T1. Increase in the percentage of embryonic weight, decrease in the percentage of Albumin and the percentage of shell at 7 days of incubation for T2 and T3. Increase in percentage of embryonic weight and amniotic sac and liquid, decrease in the percentage of Albumin and yolk, at 14 days of include sialic acid for T2. Increase the percentage of embryonic weight, and decrease the percentage of yolk at 14 days incubation for T2. They have concluded that In-Ovo injection of the hatching eggs with sialic acid contributed to increased physiological traits and embryonic development.

Key words: In-Ovo, Sialic Acid, Erythropoietin, Oxygen, Anemia, and Anoxia.

Introduction

Utilization of supplements starts on the first day after incubation when both egg albumin and yolks assist in the development of the embryo. Bird embryos develop and grow from the energy and nutrients the hen stores in the egg. Nutrients are deposited in follicles over a long period but become essential during the week before ovulation. The amount and type of nutrients the egg delivers affect how successfully an embryo develops and how healthily a chick hatches1. Sialic acids are an assortment of nine-carbon alpha-keto acid sugars. N-acetylenuraminic acid (Neu5Ac or NANA), found in animals and some prokaryotes, is the most prevalent member of this group. N-acetylgalactosamine-6-P is produced by glucosamine 6 phosphate and acetyl-CoA using a transferase to produce sialic acid. This undergoes epimerization to become N-acetylmannosamine-6-P, which then interacts with phosphoenolpyruvate to form N-acetylenuraminic-9-P (sialic acid). A monophosphate nucleoside is added to make sialic acid become cytidine monophosphate-sialic acid, which is then active to participate in the cell's oligosaccharide manufacturing process (CMP-sialic acid). The animal cell's nucleus is where this substance is made2. Sialic acid is known to serve as a red blood cell (RBC)-bound receptor for P. falciparum in humans. Also, it affords protection against oxidative stress, which may be highly important for RBCs, whose primary function is to transport oxygen3.

The development of chickens and the processes of breeding and improvement that occurred in domestic chickens during the previous years led to a change in some physiological and immunological characteristics4, the most important of which was a weakness in the membranes of Chorion, Amnion and Allantois, and thus leads to a lack of oxygen supply as a result of a decrease in the formation of cells red blood and the occurrence of a suffocation process as a result of a lack of oxygen in the body5, which is expressed by the term Anoxia in the last stages of hatching and thus the death of the embryos6. It is possible to use natural nutrients that contribute to the development of the breathing process in embryos, such as the use of sialic acid, which is the primary catalyst for the formation of the hormone erythropoie tin, which works to produce red cells through its action on the bone marrow and increases their numbers in the lining of blood vessels and increases the production of nitric oxide, which facilitates it controls blood flow, which improves oxygen supply to the heart, brain, and other tissues7.

This study aims to investigate the use of natural nutrients that contributes to solving the problem of suffocation of bird embryos and the lack of oxygen supply by increasing the production of Erythropoietin, which is one of the factors used in treating anemia in animals and humans.

Materials and methods

Animal Study

The study was conducted according to the protocol approved by the University of Anbar Ethics Committee, Iraq.

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Fertile eggs from Ross (308) strain broiler breeder hens were gotten from a commercial farm.

Experimental study

In this study, 400 eggs were Collected from Ross 308 Broiler breeders (53 weeks old). The egg was divided into 4 groups, each distributed to 100 eggs, and every one of this group was sub-divided into 4 replicates, every replicate consisting of 25 eggs. Eggs were injected using an automatic syringe that was used for oil vaccination by using a needle of 25 millimeters.

In Ovo

The eggs were injected from the broad side by making more (13mm) in the air sac than every egg injected with 100 μg sialic acid (Latex CO. Ltd, Germany) as follows: T1: The control group was placed in the incubator without injection. T2: Injected with a dose of 100 μg sialic acid at the age of zero. T3: Injected with 100 μg sialic acid dose at 7 days. T4: Injected with 100 μg sialic acid dose at 14 days of incubation. Egg candling was conducted to determine the Amnion sac for making the second injection (18 days of egg incubation), and the injection surface was sterilized with antiseptic (Dettol) before injection. The pores were closed by using Dye pedicures. Eggs were incubated in (AFLO) mark setter by distributing the groups randomly. Prepare 400 mL of sterile water divided into four glass containers. Weighed quantities of 100 mg of sialic acid and the amount of each dissolved in 100 ml of sterile water. Injection method of eggs at aged 0, 7, and 14 days of incubation, by a needle of size 25 mm, The needle is inserted from the pettition of the eggshell after piercing through the air gap depth of 13 mm and the injection dose of 100 μg of the sialic acid prepared in both dates and then underwent four tests embryonic.

Embryonic test

The first embryonic test is conducted 3 days from incubation, where we put the eggs horizontally. The shell is opened, and the following traits are measured: Embryo length, Vascular region, and Pairs of somites. The second embryonic test was conducted 7 days from incubation, where we broke the eggshell and removed the egg’s contents. The following traits are measured: Embryo weight, Albumin, and Shell. The third embryonic test was conducted 14 days after incubation, where we broke the eggshell and took out the egg’s contents; the following traits were measured: Embryo weight, yolk, amniotic sac, and liquid and Albumin. The fourth embryonic test was conducted 17 days after incubation, where we broke the eggshell and removed the egg’s contents. The following traits were measured: embryo weight and yolk.

Statistical Analysis

Complete randomization was used for this experiment (CRD). Then the SAS tool for statistical analysis was used to evaluate the data. To find significant changes between the averages, Duncan’s polynomial was used to compare the means for each treatment at significance levels of 0.05 and 0.01.

Results and discussion

The results shown in Table (1) refers to that treated the eggs with sialic acid led to a significant increase (P<0.05) in Embryo Length, Vascular length, and Pairs of somites for treatment (T2) at 3 days from incubation in the Table (2) the results refer to significant increasing (P<0.05) in Embryo weight, and significant decreasing (P<0.05) in Albumin and Shell weight for (T2) treatment compared with other treatments in 7 days of incubation. In Table (3), the results refer to a significant increase (P<0.05) in Embryo weight for (T2) treatment compared with (T0), a significant increase (P<0.05) in Amniotic Sac and Liquid compared with another treatment, significant decreasing in yolk and Albumin weight for (T2) treatment compared with another treatment.

The crucial erythropoietin response to ischemia stress is provided by the erythropoietin (EPO) regulation of red blood cell formation and its activation at low oxygen tension. Since its cloning and manufacturing, recombinant human Erythropoietin has been used clinically in anemia patients for two and a half decades, making it easier to study how

<table>
<thead>
<tr>
<th>Treatments</th>
<th>Embryo Length (Mm)</th>
<th>Vascular Region (Mm)</th>
<th>Pairs of Somites</th>
</tr>
</thead>
<tbody>
<tr>
<td>T1</td>
<td>8.00 ± 0.50b</td>
<td>9.57±0.38b</td>
<td>34.20±1.08b</td>
</tr>
<tr>
<td>T2</td>
<td>12.90±0.64a</td>
<td>12.50±0.42a</td>
<td>38.20±0.30a</td>
</tr>
<tr>
<td>T3</td>
<td>10.16±0.68a</td>
<td>11.23±0.45b</td>
<td>35.33±0.50a</td>
</tr>
<tr>
<td>T4</td>
<td>10.56±0.68a</td>
<td>10.43±0.45ab</td>
<td>35.53±0.50ab</td>
</tr>
</tbody>
</table>

* Different lowercase letters within a column indicate significant differences at the level of probability (P <0.05)
* Average ± standard error

Table 1. Effect of injection of eggs hatching in different concentrations of sialic acid in embryonic growth at the age of 3 days from incubation (%As a percentage relative to egg weight at examination).
it works14. Reports of ischemia stress in vitro and injury in animal and cell models point to possible erythropoietin benefits beyond red blood cell formation. Including vascular endothelial response to increasing nitric oxide production15 facilitates oxygen delivery to the brain16, heart and non-hematopoietic tissues. This review addresses these and other findings of erythropoietin function outside of the generation of red blood cells, including how it affects metabolism and animal models of obesity. Observations of erythropoietin activity in cell and animal model systems17, including mice with tissue-specific deletion of erythropoietin receptor (EpoR), suggest the potential for erythropoietin response in metabolism and disease17,18.

As a function of animals, Kumar and Rizvi (2013)19 note a considerable drop in RBC sialic acid content and an increase in plasma sialic acid. A reliable indicator of aging is the sialic acid content of the erythrocyte membrane, which decreases as rat age increases3. Greater expression of acute phase proteins and increased organ damage are two possibilities for the cause of elevated plasma sialic acid. Hemoglobin6, a protein that makes up 95% of red blood cells, cooperatively binds oxygen for transportation to the tissues from the lungs20. As a result, Erythropoietin’s main job is to control how much oxygen is delivered through the synthesis of red blood cells21. This job is made more accessible by the hypoxia-induced elevation of erythropoietin gene transcription, which makes erythropoietin production sensitive to the local oxygen environment22. Erythropoietin stimulates cell survival, proliferation, and differentiation by attaching to a particular receptor on the surface of erythroid progenitor cells23. Mice lacking Erythropoietin or the erythropoietin receptor have severe anemia, which causes embryonic mortality24.

Injecting the eggs with sialic acid resulted in the embryos benefiting from the acid by manufacturing erythropoietin21; since sialic acid is a significant factor in the production of the hormone erythropoietin and a crucial component of erythropoiesis, Erythropoietin has been commercialized in several forms to treat anemia in animals25. This study observed the development of the embryos of the treated injected with sialic acid. So, as mentioned above, the production of red blood cells as a result of the increase in the hormone erythropoietin led to an increase in the acceleration of biochemical processes and, thus, an increase in oxygen transfer and the ability of hemoglobin to transport it inside living cells, and thus helps to complete the process Embryo growth and development26.

### Table 2. Effect of injection of eggs hatching in different concentrations of Sialic acid in embryonic growth at the age of 7 days from incubation (%As a percentage relative to egg weight at the examination.

<table>
<thead>
<tr>
<th>Treatments</th>
<th>Embryo Weight</th>
<th>Albumin</th>
<th>Shell</th>
</tr>
</thead>
<tbody>
<tr>
<td>T1</td>
<td>1.10±0.04c</td>
<td>25.57±0.71a</td>
<td>11.97±0.33a</td>
</tr>
<tr>
<td>T2</td>
<td>3.03±0.07a</td>
<td>14.65±0.59c</td>
<td>9.31±0.35c</td>
</tr>
<tr>
<td>T3</td>
<td>2.97±0.04ab</td>
<td>17.76±0.74b</td>
<td>10.34±0.22b</td>
</tr>
<tr>
<td>T4</td>
<td>2.38±0.04b</td>
<td>17.25±0.64b</td>
<td>10.44±0.22b</td>
</tr>
</tbody>
</table>

Significant * NS NS *

Average ± standard error

### Table 3. Effect of injection of eggs hatching in different concentrations of sialic acid in embryonic growth at the age of 7 days from incubation (%As a percentage relative to egg weight at the examination.

<table>
<thead>
<tr>
<th>Treatments</th>
<th>Embryo Weight</th>
<th>Yolk</th>
<th>Amniotic Sac And Liquid</th>
<th>Albumin</th>
</tr>
</thead>
<tbody>
<tr>
<td>T1</td>
<td>22.19±0.26c</td>
<td>33.36±0.2</td>
<td>31.35±0.45</td>
<td>10.9 ± 0.23a</td>
</tr>
<tr>
<td>T2</td>
<td>29.72±0.25a</td>
<td>33.53±0.2</td>
<td>31.11±0.14</td>
<td>4.1±0.14c</td>
</tr>
<tr>
<td>T3</td>
<td>26.5±0.24 b</td>
<td>33.27±0.2</td>
<td>33.12±0.19</td>
<td>7.33±0.21b</td>
</tr>
<tr>
<td>T4</td>
<td>27.62±0.25 b</td>
<td>33.53±0.2</td>
<td>33.09±0.14</td>
<td>7.31±0.14b</td>
</tr>
</tbody>
</table>

Significant * NS NS *

Average ± standard error
In Figure 1, the results refer to a significant increase \((P<0.05)\) in Embryo Weight at the age of 17 days from incubation for \((T2)\) compared with \((T1)\). In comparison, there is a significant increase \((P<0.05)\) in Embryo Weight at the age of 17 days from incubation for \((T3 \text{ and } T4)\) compared with \(T1\). However, the results show a significant improvement \((P<0.05)\) in Yolk Weight at the age of 17 days from incubation for \((T2)\) compared with \((T1)\) and a significant improvement \((P<0.05)\) at the period of 17 days from incubation for \((T2)\) compared with another treatment. At the same time, there is a significant improvement \((P<0.05)\) in Yolk Weight at the age of 17 days from incubation for \((T3 \text{ and } T4)\) compared with \(T1\).

According to EPO, it plays a crucial role in the availability and improvement of the manufacture of red blood cells, increasing oxygen availability and thus improving the growth process\(^{27}\). The increase in oxygen in the egg means an increase in the gas exchange process between the embryo and the external environment. Thus this leads to an increase in biological processes such as metabolism and protein production in the body and obtains an increase in the number and size of muscle cells and, thus, an increase in weight for embryos\(^{28}\). At the same time, this above hypothesis can be confirmed by observing the decrease in the weight of the yolk concerning the treatments injected with sialic acid. The Chick’s embryo begins to switch its nutrition from albumen to yolk within 14 days of incubation; therefore, the indicator of measuring the yolk weight is significant. If the yolk weight decreases, this indicates the consumption of a more significant amount of it\(^{29}\), and it means that there is an increase in the feeding process for the embryo, which works to increase the weight\(^{10}\), as we indicated previously, and this is what was done in the injected experimental treatments. So from Figure 1, we notice a significant decrease in yolk weight for all treatments except \(T1\).

**Conclusions**

A problem occurs in the embryos that prevent the completion of the hatching; a shortage of oxygen occurs at the end of the incubation period. Thus, in this study, we solved this problem by injecting eggs with sialic acid and measuring some physiological and biological indicators that biologists contribute to generating an amount of oxygen that helps the embryo overcome the stressful hatching process.

**Supplementary Materials**

The following are available in this PDF, Table S1: Composition culture medium, Sheet 1 S2: Total cost, Sheet 2 S2: Stages of production, Sheet 3 S2: Direct and indirect labor, Sheet 4 S2: Culture medium, Sheet 5 S2: IMC, Sheet 6 S2: 6. Assumptions.

**Author Contributions**

Conceptualization, Ana Maria Henao Ramírez and Aura Inés Urrea Trujillo; methodology and software, Hernando David Palacio Hajduck and Ana Maria Henao Ramírez; validation and formal analysis, Ana María Henao Ramírez; investigation, resources, data curation, writing—original draft preparation, Ana María Henao Ramírez; writing—review and editing and supervision, Aura Inés Urrea Trujillo. All authors have read and agreed to the published version of the manuscript.

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Conflicts of Interest

The authors declare no conflict of interest.

Bibliographic references